32P LABELED RIBOPROBES FOR NORTHERNS

1. Add the following in order:

H ₂ 0	1.75 μl
1 mgml ⁻¹ BSA	2.5 μl
5 x buffer	5 μΙ
triton X-100	0.25 μl
1M DTT	0.5 μΙ
RNAguard	2 μΙ
10mM ATP	1 μΙ
10mM CTP	1 μΙ
10mM GTP	1 μΙ
100μM UTP	3 μΙ
DNA	1 μΙ (1 μg)
α^{32} PUTP 10mCiml $^{-1}$	5 μl
T3/T7/SP6 polymerase	1μΙ

- 2. Incubate at 30°C for 3 hours.
- 3. Dilute into 100 µl by adding:

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2 \mu l RNAGuard 10 \mu l 10 x DNAase buffer (manufacturer's) 10 \mu l 100mM DTT 52 \mu l H_2O
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Remove 2 \times 1 μl and spot onto DE81 paper for quantification later.

Add 1 μl DNAase and incubate for 15' at 37°C.

- 4. Phenol/chloroform extract.
- 5. Ethanol precipitate by adding 50 μl 7.5 M NH4OAC, 2 μl 50 mgml $^{\text{-}1}$ tRNA and 376 μl ethanol.

6. Precipitate and resuspend in 500 μl 50% formamide.

Quantification.

Rinse 1 filter: $5 \times 5'$ in 0.5 M Na₂HPO₄

 $2 \times 1'$ in dH_2O

 $2 \times 1'$ in 95% ethanol

Dry thoroughly.

Count both filters in aqueous scintillant. Washed filter represents incorporated counts.