GROWING MOSS FROM SPORES

Prepare

- Sterile pairs of forceps
- Growth media plates (see below) with cellophane discs on top.
- 1. Harvest sporogons under sterile conditions with a pair of forceps. They should look swollen and brown, and should come off easily as you pull. Handle with care as they may be ready to burst. Transfer to 900 μ l sterile dH₂O.
- 2. Add 100 μ l sodium hypochlorite, and incubate for 5 min. Invert to mix at ~1 min intervals.
- 3. Wash the sporogons with 1 ml sterile dH_2O 3-4 times. (At each wash add fresh dH_2O and invert the tube several times. As the sporogons sink to the bottom, remove the liquid).
- 4. Take out one sporogon by sucking it gently in a blue pipette tip and put it in a tube containing 200 μ l sterile dH₂O. Squash it against the side with the same tip to burst it.
- 5. Add 800 μ l sterile dH₂O and mix well by pipetting up and down.
- 6. Add 200 μl suspension to each plate, and spread it out by adding 400-1000 μl dH2O to it.
- 7. Incubate at 25 °C in continuous light.

Growth media for spore germination

5 ml solution B

5 ml solution C

5 ml solution D

5 ml 500mM ammonium tartrate

4g agar

dH₂O to make up to 490 ml

Autoclave

Add 10 ml sterile 500mM CaCl2

Mix and pour plates.

Stock solution B:

 $MgSO_4.7H_2O$ (magnesium sulphate 7-hydrate) 2.5 g (or 1.2 g of anhydrous $MgSO_4$)

 dH_2O to 100 ml

Make several 2.5 ml aliquots and store these and any remaining solution at -20°C.

Stock solution C:

 KH_2PO_4 (potassium phosphate) 2.5 g dH_2O to 50 ml

Adjust pH to 6.5 with minimal volume of 4 M KOH, then make up to 100 ml with additional dH₂O. Make 2.5 ml aliquots (as above) and store at $-20^{\circ}C$.

Stock solution D:

 KNO_3 (potassium nitrate) 10.1 g FeSO₄.7H₂O (iron sulphate 7-hydrate) 0.125 g dH₂O to 100 ml

Make aliquots and store at $-20^{\circ}C$ (as above).